

Mass Spec data available on most of the websites, not yet available for MicrosporidiaDB or PiroplasmaDB

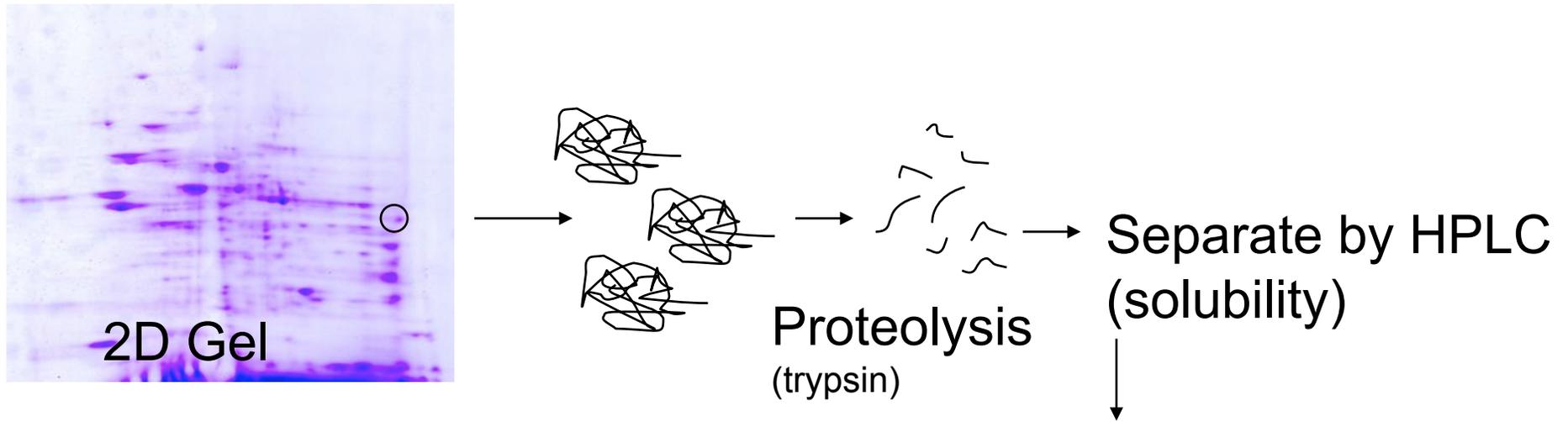
Mass Spectrometry at EuPathDB

- AmoebaDB
- CryptoDB
- GiardiaDB
- MicrosporidiaDB
- PiroplasmaDB
- PlasmoDB
- ToxoDB
- TrichDB
- TriTrypDB

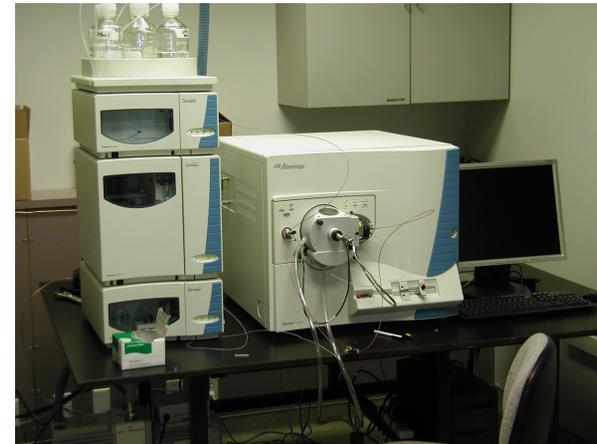
MS Use Cases

- evidence for protein expression
- evidence for subcellular localization
- confirm gene structural annotation
- detect missed gene annotations

Tandem Mass Spectrometry (MS/MS)



One example MS methodology for protein analysis is presented. There are many other techniques that vary in sample preparation and instruments used.



1. Separate by peptide mass (MS)
2. Fragment, analyze component masses (MS)

ORFs with Mass Spectrometry Evidence

- CryptoDB
- GiardiaDB
- ToxoDB

The screenshot displays the CryptoDB website interface. At the top, the logo for CryptoDB (Cryptosporidium Genome Resource) is visible, along with the version number 4.1 and the date June 2009. A search bar contains the gene ID 'cgd7_230' and a text search for 'membrane'. Navigation tabs include Home, New Search, My Searches (3), Tools, Data Sources, Downloads, and Community. A news section on the left lists updates from February 2009 to June 2007. The main content area features three columns: 'Identify Genes by:' with options like Text, IDs, Species, Genomic Position, Gene Attributes, Protein Attributes, Protein Features, SimilarityPattern, Transcript Expression, Protein Expression, Cellular Location, Putative Function, Evolution, and Population Biology; 'Identify Other Data Types:' with options like Isolates, Genomic Sequences, SNPs, ESTs, Assemblies, ORFs (with a sub-menu for Orf ID(s), Mass Spec. Evidence, Protein Motif Search, BLAST, and Chromosomal Location); and 'Tools:' including BLAST, Sequence Retrieval, PubMed and Entrez, GBrowse, and CryptoCyc. The footer contains the website URL, copyright information, a contact prompt, and the EuPathDB logo.

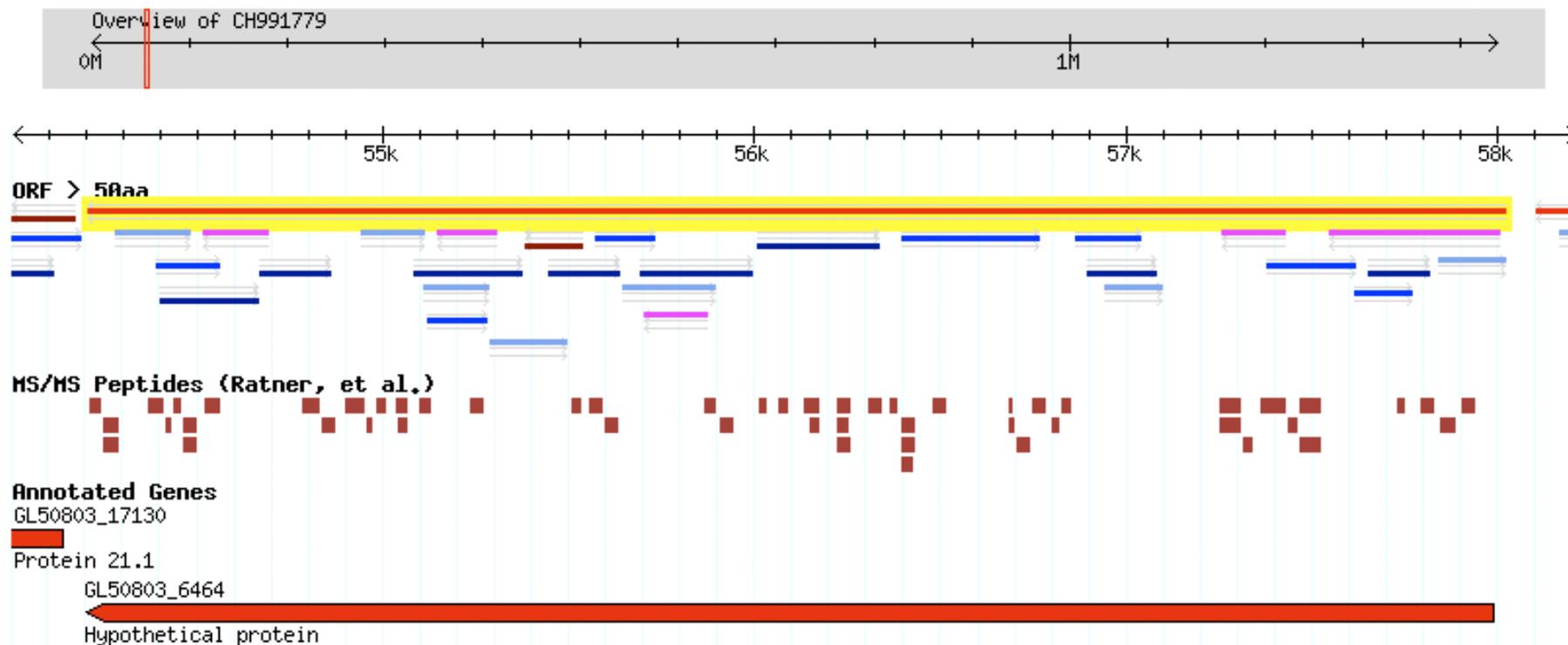
ORFs at EuPathDB

AC**TGA**TCCAGCTCAGTACATTTTAGATACTTCTATTTATTCATCTGG**TGA**TTTA

S S S V H F R Y F Y L F I W

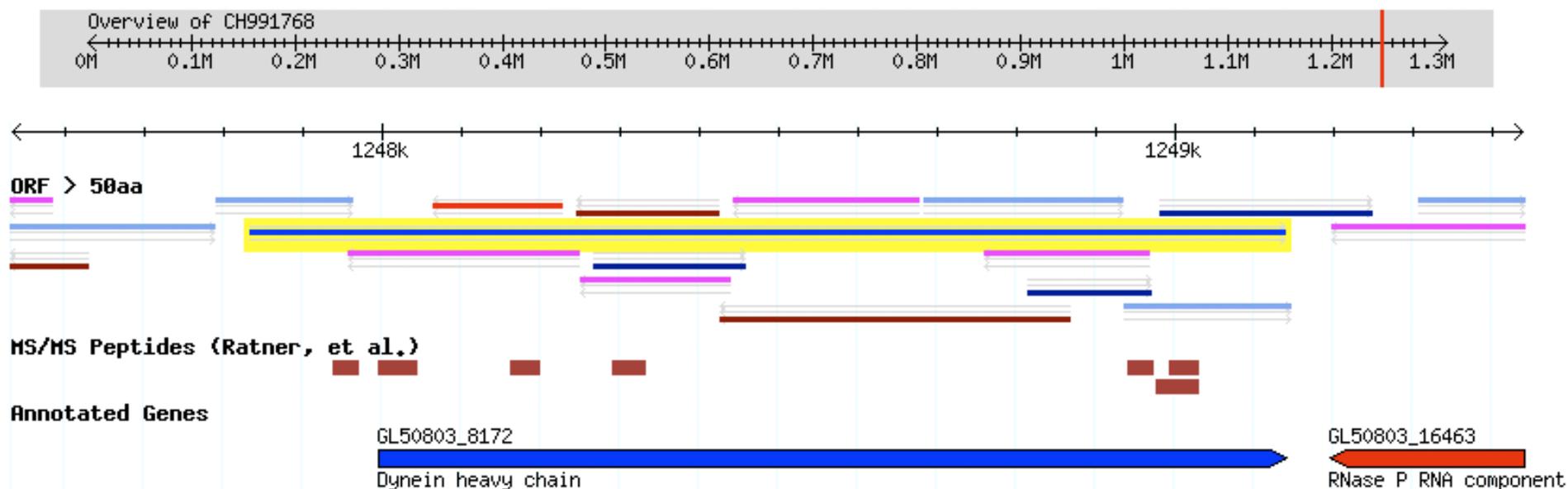
- computed in all 6 reading frames
- do not necessarily begin with Met
- are not annotated or reviewed

Re-Mapping Peptide ORF Hits to Genes



MS analysis often includes peptides identified by spectra searches against an ORF database. Frequently those ORFs correspond to annotated genes, as shown here. Such ORFs are typically not very interesting to our community. So, in these cases we move the MS identified peptides from the ORF to the annotated protein. In the example shown here, the peptides shown were originally assigned, by the data provider, to the ORF highlighted in yellow. We moved the peptides to the GL50803_6464 gene. A search for 'genes with mass spec evidence' will return the annotated gene. A search for 'orfs with mass spec evidence' will NOT return this ORF because we have taken the MS evidence away from the ORF and put it on the gene.

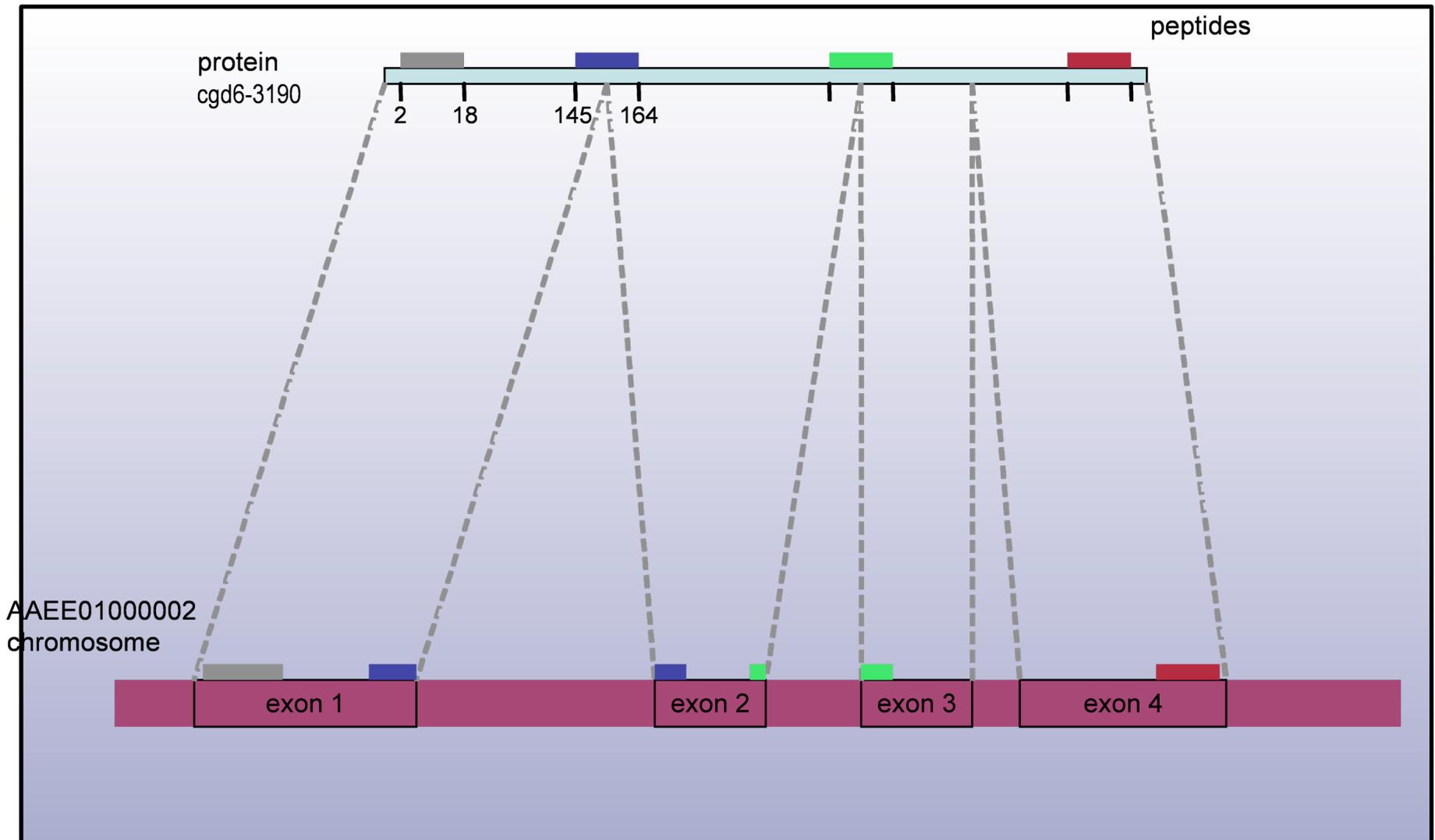
Re-Mapping ORF Peptide Hits to Genes or Not



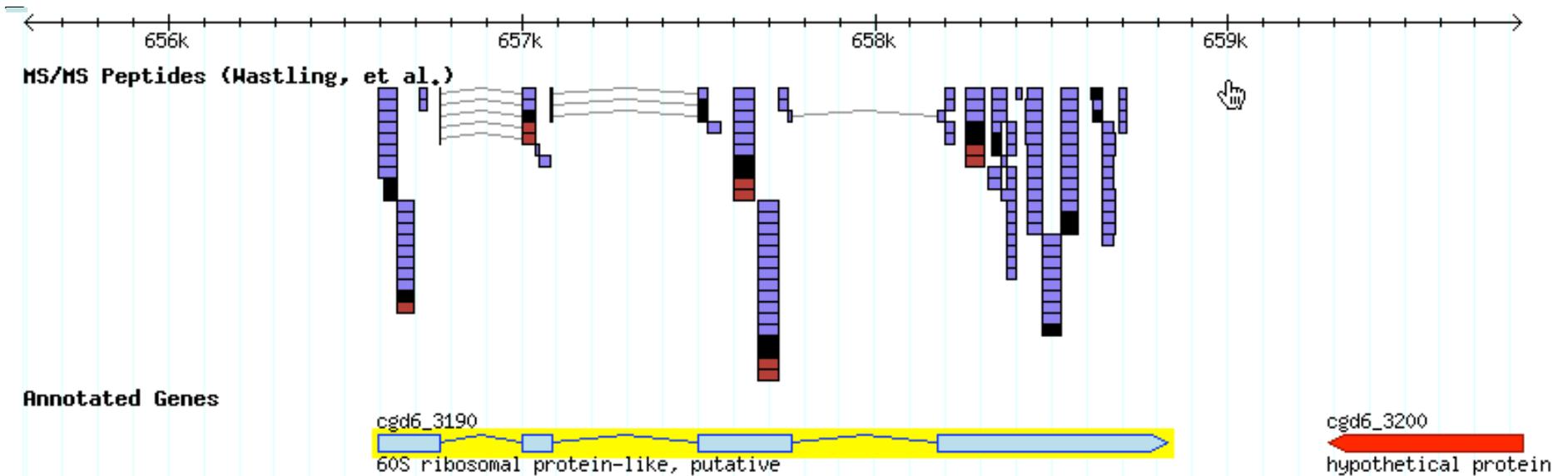
In the example shown here, the peptides shown were originally assigned, by the data provider, to the ORF highlighted in yellow. Because one of the peptides identified on the ORF does not correspond to the GL5083_8172 gene, we leave all the assigned peptides associated with the original ORF. A search for ORFs with MS evidence WILL return this ORF. By moving the peptides from ORFs that are fully contained within an annotated gene (previous slide), the search for possibly interesting identifications, like this example, is made easier. However, be very cautious about attempting to reannotate a gene on the sole basis of a single MS peptide, as shown here. That one peptide may have been mis-identified by the MS software.

The data we get from the research labs includes the peptide coordinates on the protein sequence. We are able to calculate the corresponding genomic coordinates for the DNA sequence that encodes the peptide. In some cases those sequences span an intron. Because the gene exons have to be spliced to generate the detected peptide sequences, such examples support the annotated intron/exon boundaries.

Map Peptides to Genome



Support for Intron Boundaries



some examples where the DNA sequences encoding an MS peptide span an intron. This support the intron/exon boundaries annotated correctly.

Instructions

Search using a sequence name, gene name, locus, or other landmark. The wildcard character * is allowed. To center on a location, click the ruler. Use the Scroll/Zoom buttons to change magnification and position.

Examples: [AAEE0100001:1-35000](#), [AAEE0100002:106357-116356](#), [AAEL0100026:7652-27651](#), [cgd6_780](#), [CMU_011540](#).

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Search

Landmark or Region:

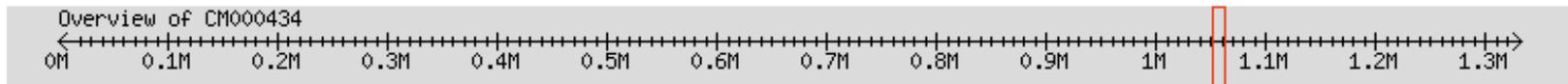
Reports & Analysis:

Data Source

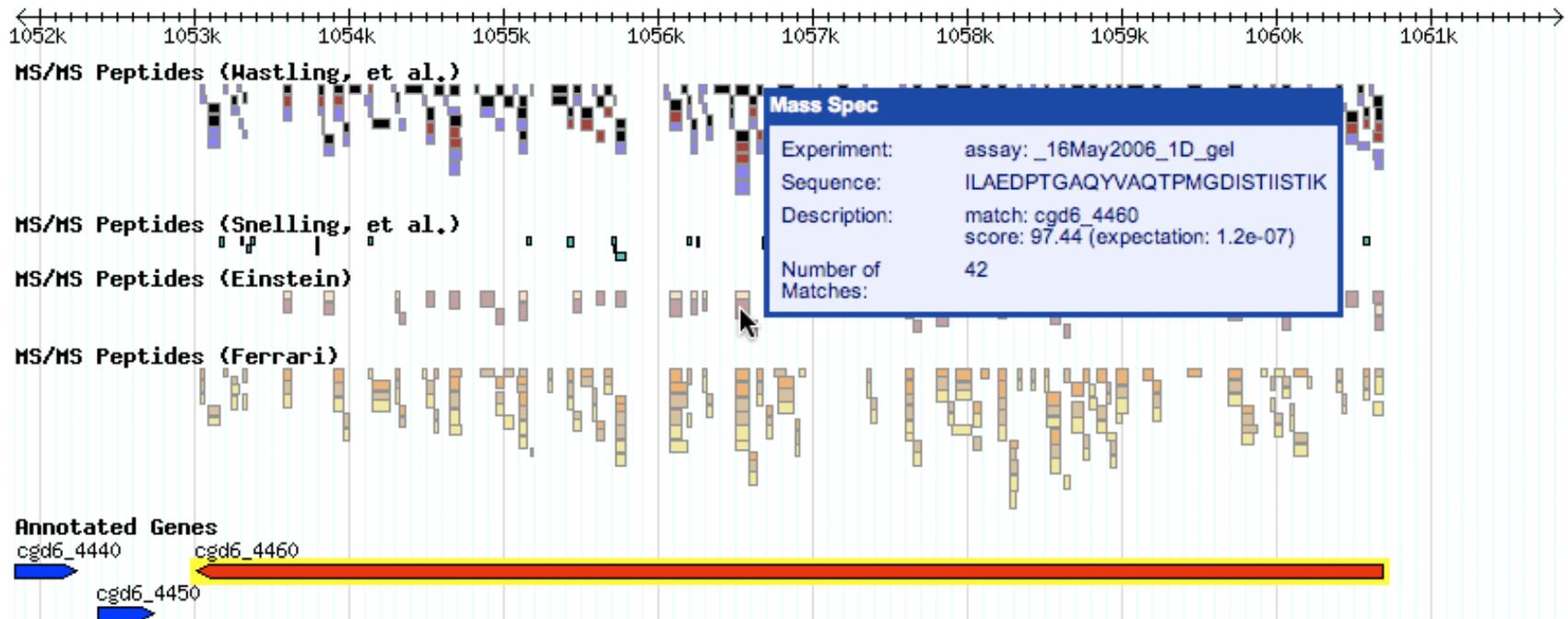
Scroll/Zoom: Flip

Select the Data Source of your choice (above).

Overview



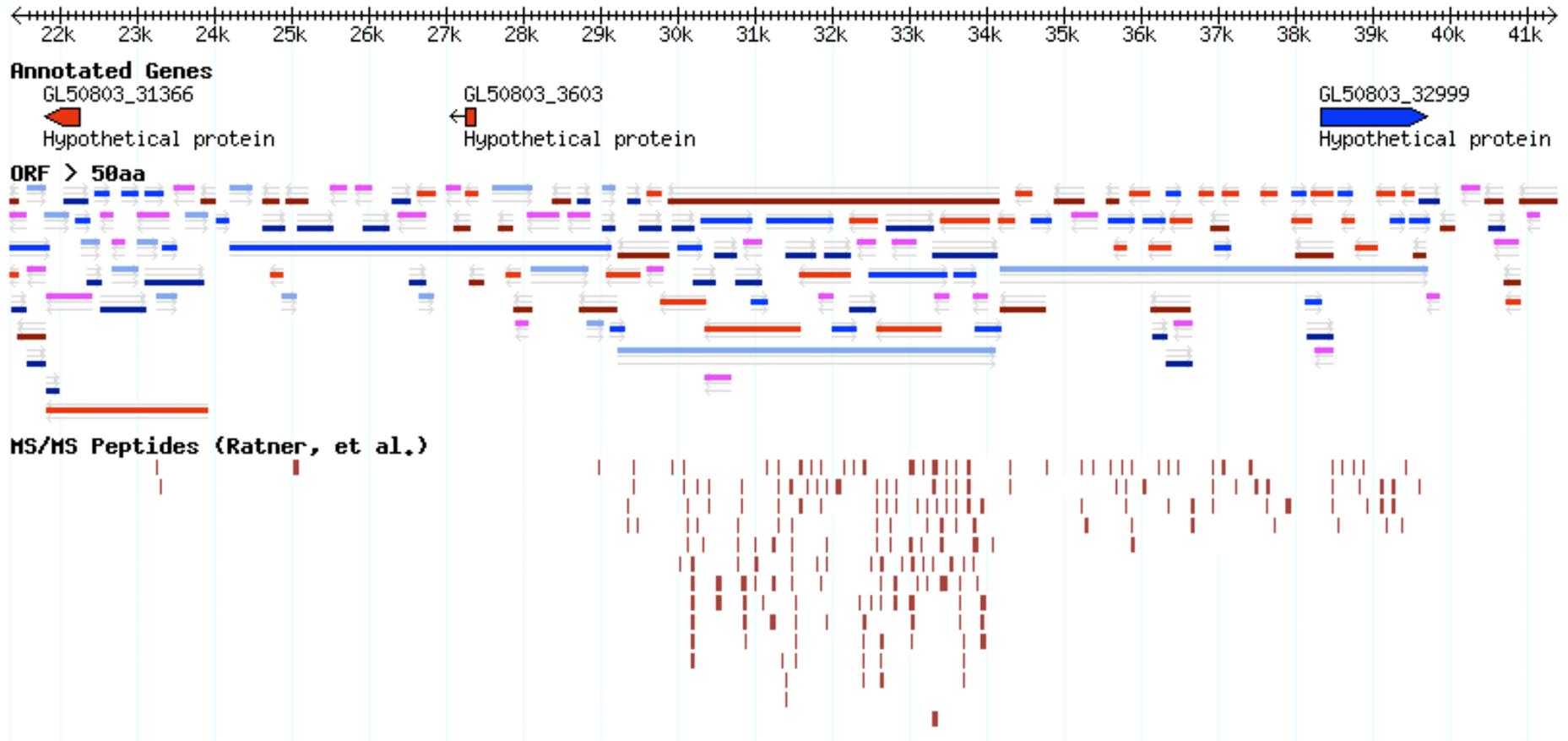
Details



[Clear highlighting](#)

Genome Browser view. This is similar to what was seen on the gene record page but here you have more control over which tracks and specific genome sequence range are shown. To keep the display reasonably compact, we do not display redundant peptide sequences within a given experiment. Instead we show a single block representing the peptide and in a mouseover popup we report the "Number of Matches" seen for that sequence.

Reveal Missing Gene Annotations



This is a region of a Giardia chromosome where long open reading exist but there is no gene annotated or the annotated gene does not extend for the length of the ORF. There were many peptides identified for this region by searching spectra against the ORF sequences suggesting that the associated ORFs are part of a gene that has not yet been annotated.