

Phenotypic data

Learning objectives:

- Explore how to combine different phenotypic data
- Explore high throughput mutagenesis data
- Explore curated phenotypic data
- Explore high throughput subcellular localization data

1. Identify genes that are targeted to the ciliary tip of *Trypanosoma brucei* that are also essential for parasite fitness.

Note for this exercise use <http://tritrypdb.org>

- TriTrypDB integrates data from the TrypTag project (<http://tryptag.org>). Genes from *T. brucei* were N- and C-terminally tagged with a fluorescent protein and subcellular localization determined by microscopy. The description of the localization was done using gene ontology terms.

- Start by finding the “Cellular Localization Imaging” search.

Identify Genes based on Cellular Localization Imaging

Search for...

cellu|

Genes

Protein targeting and localization

Cellular Localization Imaging

Reset values

Organism

Trypanosoma brucei brucei TREU927

Location of tag

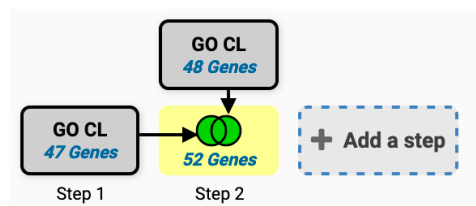
N-terminal

C-terminal

GO Term or GO ID

GO:0097542 : ciliary tip : 3

- Configure the search to identify the GO term “Ciliary Tip” – notice that when you start typing the autocomplete function offers you selectable options.
- Since the experiment examined both N and C terminal fusions proteins, you will have to run the search twice and combine the results from both searches. Did you use a union or an intersect to combine the results?



- Explore the results you got. Scroll down to the results section, then scroll to the right of the results window to reveal the subcellular localization

Product Description	# Transcripts	EC numbers	Cellular localization images
ain-like protein 1.1	1	3.4.22.17 (Transferred entry: 3.4.22.52 and 3.4.22.53)	
N repeat, putative	1	N/A	
thetical protein, erved	1	N/A	
thetical protein, erved	1	N/A	
0-like serine/threonine-kinase, putative	1	N/A	
thetical protein,	1	N/A	

images. These are very small, but you can right click on them to open a larger image in a new window.

- b. Add a step to identify how many genes are essential for the fitness of the parasite. Click on Add step, then search for the phenotype searches. Click on the Phenotype Evidence option.

Combine with other Genes

1 Choose *how* to combine with other Genes

2 INTERSECT 3 2 UNION 3 2 MINUS 3 3 MINUS 2

2 Choose *which* Genes to combine. From...

A new search An existing strategy My basket

phen
Phenotype
 Phenotype Evidence

- Select the “High-throughput phenotyping using RNAi target sequencing (David Horn)”.

Add a step to your search strategy

Search for Genes by Phenotype Evidence

The results will be intersected with | v the results of Step 2.

Filter Data Sets:

Legend: CP Curated Phenotype PQ Quantitative Phenotype PT Phenotype Text

Organism	Data Set	Choose a Search
<i>Trypanosoma brucei</i> /brucei TREU927	<input checked="" type="checkbox"/> High-throughput phenotyping using RNAi target sequencing (David Horn)	<input checked="" type="checkbox"/> PQ
<i>Trypanosoma brucei</i> /brucei TREU927	<input type="checkbox"/> Sanger siRNA Phenotypes (Sanger)	<input type="checkbox"/> CP <input type="checkbox"/> PT

- Configure the search to return genes that are decreased in coverage by 1.5 fold when comparing the maximum expression value of all induced samples to the uninduced sample.

For the Experiment

- Quantitated from the CDS Sequence
- Quantitated from gene model (5 prime UTR + CDS)

select all | clear all

return protein coding Genes

that are Decrease in coverage

with a Fold change >= 1.5

between each gene's maximum expression value

in the following Reference Samples

- Uninduced sample

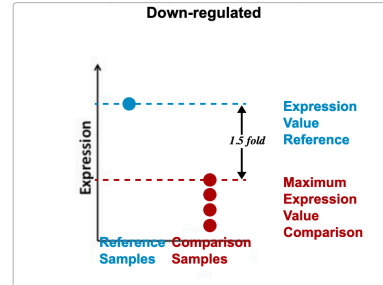
select all | clear all

and in the following Comparison Samples

- Induced in bloodstream (BS) forms, 3 days (10 doublings)
- Induced in bloodstream (BS) forms, 6 days (20 doublings)
- Induced in procyclic forms (PS) forms, 9 days (9 doublings)
- Induced throughout differentiation (DIF = 7 BS doublings + 6 PS doublings)

select all | clear all

Example showing one gene that would meet search criteria
(Dots represent this gene's expression values for selected samples)



For each gene, the search calculates:

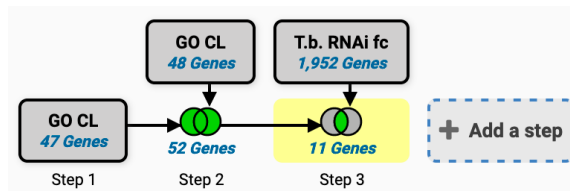
$$\text{fold change} = \frac{\text{reference expression value}}{\text{maximum expression value in comparison}}$$

and returns genes when fold change >= 1.5.

You are searching for genes that are down-regulated between one reference sample and at least two comparison samples.

This calculation creates the narrowest window of expression values in which to look for genes that meet your fold change cutoff. To broaden the window, use the average or minimum comparison value.

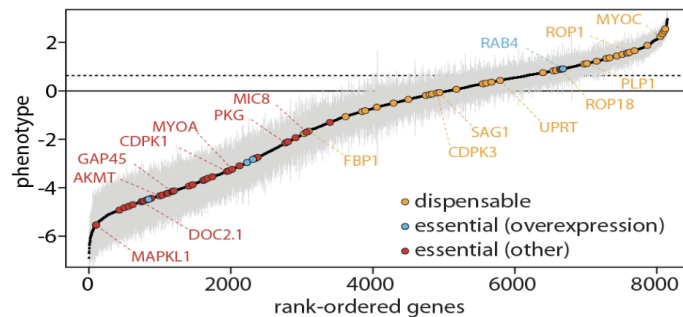
- How many genes did you get?



2. Finding genes based on high throughput mutagenesis and fitness analysis.

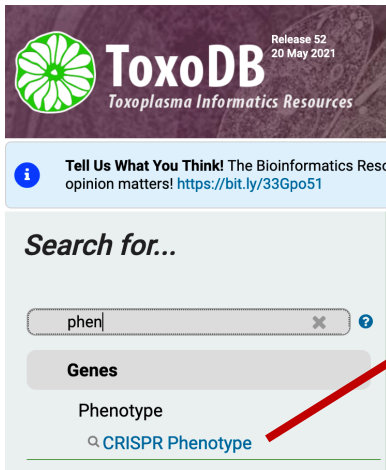
Note for this exercise use <http://toxodb.org>

- Navigate to the CRISPR phenotype search. Note that this search form is quite simple just requiring a range of fitness values. The defaults return all genes not limiting the search at all. This is only useful in as much as it tells you which genes were assayed which is nearly the entire genome. The tricky bit is deciding where to make the cutoffs. Again, the description on the search form is very helpful in this regard



(as is the link to the paper ... remember these phenotypes were assayed under specific conditions so just because a particular gene doesn't show a phenotype doesn't mean it wouldn't in other conditions (or infecting an

actual host). The plot showing the phenotype score (fitness) is particularly useful. Red points along the plot are genes known to be essential under these conditions while yellow are known to be expendable. This will help you determine where to set the values. The scores range from 2.96 (least “essential) to -6.89 (most “essential). Try it running this search by limiting the range from -6.89 to -4. Do you get the expected results based on the above graph and the number of genes returned in your search results?



Identify Genes based on CRISPR Phenotype

Phenotype Score >=

-6.89

Phenotype Score <=

-4

CRISPR
1,343 Genes

+ Add a step

Step 1

- What kinds of genes are in your results? What kinds of genes would you expect to be essential? One way to explore the data is to run a GO enrichment analysis to determine if any biological processes are enriched in your results. Give this a try. What do your results look like and do they make sense?

Gene Results | Genome View | Gene Ontology Enrichment x | Metabolic Pathway Enrichment x | Analyze Results

[Rename This Analysis | Duplicate]

Gene Ontology Enrichment

Find Gene Ontology terms that are enriched in your gene result. [Read More](#)

Parameters

Organism: Toxoplasma gondii GT1

Ontology: Cellular Component, Molecular Function, Biological Process

Evidence: Computed, Curated

Limit to GO Slim terms: No, Yes

P-Value cutoff: 0.05 (0 - 1)

Submit

Analysis Results: 243 rows

GO ID	GO Term	Genes in the bkgd with this term	Genes in your result with this term	Percent of bkgd genes in your result	Fold enrichment	Odds ratio	P-value	Benjam
GO:0010467	gene expression	493	235	47.7	2.35	4.38	7.07e-48	6.50e-45
GO:0034645	cellular macromolecule biosynthetic process	385	194	50.4	2.49	4.72	1.82e-43	8.36e-41

- How many of these genes are upregulated in *in vivo* chronic stages of *T. gondii*?
- Click on add step and elect the RNAseq searches under the Transcriptomics category

- Find the experiment with chronic stages and run a search based on differentially expressed genes (DE).

The results will be intersected with the results of Step 1.

Filter Data Sets: Legend: Differential Expression Fold Change Percentile SenseAntisense

Organism	Data Set	Choose a Search
Toxoplasma gondii ME49	Transcriptome during acute or chronic infection in mouse brain (Pittman et al.)	<input checked="" type="button" value="DE"/> <input type="button" value="FC"/> <input type="button" value="P"/>

- Intersect genes that are 2-fold upregulated in chronic stages compared to acute stages.

← Add a step to your search strategy ⓘ

Experiment

Acute and chronic *T.gondii* infection of mouse. unstranded

Reference Sample

acute infection 10 days p.i.
 chronic infection 28 days p.i.

Comparator Sample

acute infection 10 days p.i.
 chronic infection 28 days p.i.

Direction

up-regulated

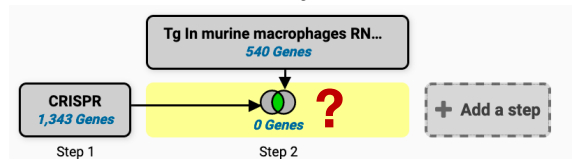
fold difference >=

2

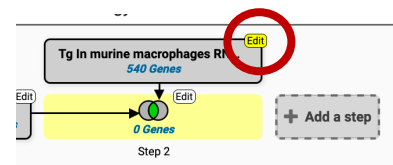
adjusted P value less than or equal to

0.1

- Did you get zero results? This is to be expected since the CRISPR data was analyzed using the GT1 strain of *Toxoplasma* and the RNA-Seq data is from the ME49 strain. How can you fix this?



- Hint: transform the results in step 2 from *T. gondii* ME49 to *T. gondii* GT1. Click on the step edit button (move your mouse over the step and select edit).



- Select **orthologs** from the menu items at the top of the pop window.

View | Analyze | Revise | Make nested strategy | Insert step before | **Orthologs** | Delete ×

Details for step *Tg In murine macrophages RNA-Seq (de)* ↗
540 Genes

Experiment Acute and chronic T.gondii infection of mouse. unstranded

Reference Sample acute infection 10 days p.i.

Comparator Sample chronic infection 28 days p.i.

Direction up-regulated

fold difference >= 2

adjusted P value less than or equal to 0.1

▶ Give this search a weight

- Select *T. gondii* GT1 from the list of organisms and click on Run Step.

Organism

1 selected, out of 31

[add these](#) | [clear these](#) | [select only these](#)
[select all](#) | [clear all](#)

gt1 × ?

Sarcocystidae

Toxoplasma

Toxoplasma gondii GT1

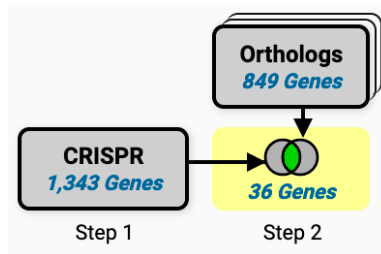
[add these](#) | [clear these](#) | [select only these](#)
[select all](#) | [clear all](#)

Syntenic Orthologs Only?

no ?

Run Step

- Now what do your results look like?



3. Identify essential *Plasmodium falciparum* genes that are highly expressed in schizont stages of the parasite.

Note for this exercise use <https://plasmodb.org>

- You can start by exploring the phenotype data in PlasmoDB.
- Select and run the search associated with the dataset: piggyBac insertion mutagenesis (John Adams).

Search for...

phen

Genes

Phenotype

Phenotype Evidence

Identify Genes based on Phenotype Evidence

Filter Data Set: Legend: Association to Genomic Segments Curated Phenotype Similarity Similarity of Association Phenotype Text

Organism	Data Set	Choose a Search
<i>Plasmodium berghei</i> ANKA	P. berghei knockout (PlasmoDEM) growth phenotypes (Bushell, Gomes and Sanderson et al.)	<input type="checkbox"/>
<i>Plasmodium berghei</i> ANKA <i>Plasmodium falciparum</i> 3D7 <i>Plasmodium yoelii</i> yoelii 17XNL	RMgmDB - Rodent Malaria genetically modified Parasites (Chris J. Janse)	<input type="checkbox"/>
<i>Plasmodium falciparum</i> 3D7	eDTL for HB3, Dd2 and 34 progeny (Gonzales et al.)	<input type="checkbox"/>
<i>Plasmodium falciparum</i> 3D7	piggyBac insertion mutagenesis (John Adams)	<input checked="" type="checkbox"/>

- Configure the search to identify genes with a *mutant fitness score* of less than -3. Note that you can select the range by either clicking and dragging your mouse over the histogram or by typing the values in the selection boxes.

Identify Genes based on piggyBac insertion mutagenesis (mutant fitness and mutagenesis index scores)

Reset values

Genes

5,385 Genes Total

expand all | collapse all

Find a variable

[Mutagenesis Index Score](#)

[Mutant Fitness Score](#)

856 of 5,385 Genes selected **Mutant Fitness Score**

Min: -4.09 Mean: -2.25 Median: -2.68 Max: 2.77

Select Mutant Fitness Score from to

5,385 (100%) of 5,385 Genes have data for this variable

- How many genes did you identify? Which gene has the lowest fitness score? Note that you might need to add the fitness score column, by clicking on add columns then filtering the options with the word “fitness”.

The screenshot shows the 'Select Columns' dialog box in PlasmoDB. The search field contains 'fitn'. The list of options includes 'P.falciparum 3D7 piggyBac insertion mutagenesis - mutant fitness score'. A red circle highlights the search field and the selected option. A red arrow points from the 'Add Columns' button in the background to the 'Update Columns' button in the dialog.

Gene ID	Transcript	Location	Product Description	Score
PF3D7_0914400	PF3D7_0914400.1	Plasmodium falciparum 3D7	protein KIC3	-4.094
PF3D7_1144100	PF3D7_1144100.1	Plasmodium falciparum 3D7	mitochondrial large subunit ribosomal protein, putative	-4.036
PF3D7_0728400	PF3D7_0728400.1	Plasmodium falciparum 3D7	SDH5 domain-containing protein, putative	-4.024

- Click on Add Step and find the RNA-Seq searches.

The screenshot shows the 'Add a step to your search strategy' dialog. The 'Combine with other Genes' option is selected. The 'Choose which Genes to combine. From...' section shows a search field with 'rna' and a list of options including 'RNA-Seq Evidence'. A red arrow points from the 'Add a step' button in the background to the 'Combine with other Genes' option.

- Find the search called “Intraerythrocytic development cycle transcriptome (2019)” and select the percentile search.

Search for Genes by RNA-Seq Evidence

The results will be intersects with the results of Step 2.

Filter Data Sets: Legend: DE Differential Expression FC Fold Change P Percentile SA SenseAntisense

Organism	Data Set	Choose a Search
<i>Plasmodium falciparum</i> 3D7	Intraerythrocytic development cycle transcriptome (2019) (Wichers et al. 2019)	DE FC P SA
<i>Plasmodium falciparum</i> 3D7	Intraerythrocytic development cycle transcriptome (2018) (Toenhake et al.)	FC P SA
<i>Plasmodium falciparum</i> 3D7	Transcriptome during intraerythrocytic development (Bartfai et al.)	FC P
<i>Plasmodium falciparum</i> 3D7	Blood stage transcriptome (3D7) (Otto et al.)	FC P
<i>Plasmodium falciparum</i> 3D7	Intraerythrocytic cycle transcriptome (3D7) (Hoeijmakers et al.)	FC P SA
<i>Plasmodium falciparum</i> 3D7	Strand specific transcriptome of the intraerythrocytic developmental cycle (Siegel et al.)	FC P SA
<i>Plasmodium vivax</i> P01	Transcription profile of intraerythrocytic cycle (Zhu et al.)	FC P

- Configure the search to identify all genes that are in the 80-100 percentile in all three available schizont samples. Remember to change the parameter to require matching all samples.
- How many genes did you get? Are any of these genes interesting? How many are predicted to be secreted?

Samples

- young ring 8 hpi
- late ring_early trophozoite 16 hpi
- mid trophozoite 24 hpi
- late trophozoite 32 hpi
- early schizont 40 hpi
- schizont 44 hpi
- late schizont 48 hpi
- purified merozoites 0 hpi

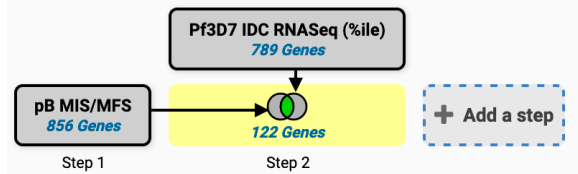
[select all](#) | [clear all](#)

Minimum expression percentile

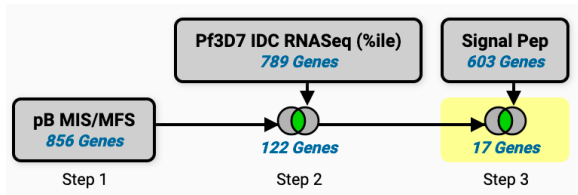
Maximum expression percentile

Matches Any or All Selected Samples?

all



- How did you identify the secreted genes? Hint, add a step and search for genes that have a predicted secretory signal peptide.



4. Identify *Neurospora crassa* genes that affect conidia formation.

Note for the exercise use <https://fungidb.org>

- Start by locating the phenotype searches.

FungiDB Release 52
20 May 2021
Fungal & Oomycete Informatics Resources

Search for...

pheno

Genes

Phenotype

Phenotype Evidence

<i>Fusarium oxysporum</i> f. sp. melonis 26406		
<i>Fusarium verticillioides</i> 7650		
<i>Heliophoma capsulatum</i> D186AR		
<i>Hyaloperonospora arabidopsidis</i> Emoy2		
<i>Metschnikowia lericii</i> populina RBAG31		
<i>Phytophthora infestans</i> T30-4		
<i>Phytophthora sojae</i> strain P6497		
<i>Puccinia graminis</i> f. sp. tritici ORL 7536-70D-3		
<i>Pyricularia oryzae</i> 70-15		
<i>Rhizopus delemar</i> RA-99-080		
<i>Saccharomyces cerevisiae</i> S288C		
<i>Sclerotinia sclerotiorum</i> 1980 UP-70		
<i>Tochodoma virescens</i> DVC2-8		
<i>Ustilago maydis</i> 521		
<i>Aspergillus fumigatus</i> AF293	Manually Curated Aspergillus Phenotypes (VEuPathDB)	CP
<i>Aspergillus nidulans</i> FGSC A4		
<i>Aspergillus niger</i> CBS 513.88		
<i>Aspergillus oryzae</i> RIB40		
<i>Cryptococcus gattii</i> WM276	Manually Curated Cryptococcus Phenotypes (VEuPathDB)	CP
<i>Cryptococcus neoformans</i> var. grubii H99		
<i>Cryptococcus neoformans</i> var. neoformans JEC21		
<i>Fusarium graminearum</i> PH-1	Manually Curated Fusarium Phenotypes (VEuPathDB)	CP
<i>Neurospora crassa</i> OR74A	Neurospora Genome Project Phenotype Image Collection (Dunlap et al.)	CP
<i>Neurospora crassa</i> OR74A	Phenotypic analysis of <i>Neurospora crassa</i> knockout mutants (Borkovich et al.)	CP
<i>Pyricularia oryzae</i> 70-15	Manually Curated Pyricularia Phenotypes (VEuPathDB)	CP

- This search provides you the option to filter based on categories on the left. Notice how when you select a different category on the left the filtering options in the middle change. Select the **Conidia number** category. Next select the “Reduced” value.

Curated Phenotype

Identify Genes based on Knockout Mutants

Reset values

Genes

1,283 Genes Total

99 of 1,283 Genes selected

Conidia Number

Keep checked values at top

1,283 (100%) of 1,283 Genes have data for this variable

Conidia Number	Remaining Genes	Genes	Distribution	%
<input type="checkbox"/> Increased	12 (1%)	12 (1%)		(100%)
<input type="checkbox"/> Normal	1,154 (90%)	1,154 (90%)		(100%)
<input type="checkbox"/> Not Formed	1 (< 1%)	1 (< 1%)		(100%)
<input type="checkbox"/> Not formed	11 (1%)	11 (1%)		(100%)
<input checked="" type="checkbox"/> Reduced	99 (8%)	99 (8%)		(100%)
<input type="checkbox"/> Severely reduced	3 (< 1%)	3 (< 1%)		(100%)
<input type="checkbox"/> Not specified	4 (< 1%)	4 (< 1%)		(100%)

- Notice that this search allows you to explore your results even before you click on the “Get Answer” button! Click around on the other categories on the left and see if the genes that are involved in a reduced number of conidia may also be involved in other phenotypes. For example, click on the **Ascospore Number** category, how maybe of your genes also have a phenotype with no ascospore formation?

Genes

1,283 Genes Total

expand all | collapse all

Find a variable

- Aerial Hyphae Height
- Ascospore Morphology
- Ascospore Number**
- Basal Hyphae Growth Rate
- Conidia Morphology
- Conidia Number
- Perithecia Morphology
- Perithecia Number
- Protoperithecia Number
- Protoperithecial Morphology

99 of 1,283 Genes selected Conidia Number X

Ascospore Number

Check items below to apply this filter

1,283 (100%) of 1,283 Genes have data for this variable

	Remaining Genes	Genes	Distribution	%
<input type="checkbox"/> Ascospore Number	99 (100%)	1,283 (100%)		
<input type="checkbox"/> Normal	32 (32%)	1,043 (81%)		(3%)
<input type="checkbox"/> Not formed	56 (57%)	169 (13%)		(33%)
<input type="checkbox"/> Reduced	11 (11%)	65 (5%)		(17%)
<input type="checkbox"/> Increased	0 (0%)	2 (< 1%)		(0%)
<input type="checkbox"/> Severely Reduced	0 (0%)	5 (< 1%)		(0%)
<input type="checkbox"/> Severely reduced	0 (0%)	1 (< 1%)		(0%)

- Click on get answer. What kinds of genes are in your results? Try analysing the results to see if there are any biological processes enriched in your results.

99 Genes (98 ortholog groups) Revise this search

Gene Results Genome View Gene Ontology Enrichment x Analyze Results

Gene Ontology Enrichment

Find Gene Ontology terms that are enriched in your gene result. Read More

Parameters

Organism: Neurospora crassa OR74A

Ontology:

- Biological Process
- Cellular Component
- Molecular Function

Evidence:

- Computed
- Curated

Limit to GO Slim terms: No

P-Value cutoff: 0.05 (0 - 1)

Submit

Analysis Results:

361 rows

Open in Revigo Show Word Cloud Download

GO ID	GO Term	Genes in the bkgd with this term	Genes in your result with this term	Percent of bkgd genes in your result	Fold enrichment	Odds ratio	P-value	B
GO:0070787	conidiophore development	84	26	31.0	22.87	44.43	1.32e-29	1.28e-
GO:0032501	multicellular organismal process	194	33	17.0	12.57	22.24	2.22e-28	1.08e-
GO:0061458	reproductive system development	184	32	17.4	12.85	22.51	8.32e-28	1.61e-
GO:0048608	reproductive structure development	184	32	17.4	12.85	22.51	8.32e-28	1.61e-
GO:0075259	spore-bearing structure development	184	32	17.4	12.85	22.51	8.32e-28	1.61e-
GO:0048731	system development	185	32	17.3	12.78	22.36	9.97e-28	1.61e-
GO:0007275	multicellular organism development	187	32	17.1	12.64	22.07	1.43e-27	1.98e-